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APPLICATION NO.	F	ILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO
09/818,086		03/26/2001	Dale Baskin	7414.0043	2844
22852	7590	12/03/2004		EXAMINER	
FINNEGA	N, HENE	DERSON, FARABO	TUNG, JOYCE		
LLP 1300 I STRI	EET NW		ART UNIT	PAPER NUMBER	
WASHING	,	20005	1637		

DATE MAILED: 12/03/2004

Please find below and/or attached an Office communication concerning this application or proceeding.

		Amplica	tion No	Applicant(s)				
Office Action Summary			ition No.	9				
		09/818		BASKIN ET AL.				
	Office Action Summary	Examir		Art Unit				
		Joyce		1637				
Period for I	The MAILING DATE of this commur Reply	ncation appears on t	me cover sneet with the	correspondence address				
A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) FROM THE MAILING DATE OF THIS COMMUNICATION. - Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication. - If the period for reply specified above is less than thirty (30) days, a reply within the statutory minimum of thirty (30) days will be considered timely. - If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication. - Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).								
Status								
1)⊠ R	esponsive to communication(s) file	ed on <u>05 Septembe</u>	<u>r 2004</u> .					
,		2b)☐ This action is						
	,—							
cl	closed in accordance with the practice under Ex parte Quayle, 1935 C.D. 11, 453 O.G. 213.							
Disposition	n of Claims							
 4) Claim(s) 1-68 is/are pending in the application. 4a) Of the above claim(s) 51-67 is/are withdrawn from consideration. 5) Claim(s) is/are allowed. 6) Claim(s) 1-50 and 68 is/are rejected. 7) Claim(s) is/are objected to. 8) Claim(s) are subject to restriction and/or election requirement. 								
Application	n Papers							
9) The specification is objected to by the Examiner.								
10) The drawing(s) filed on is/are: a) accepted or b) objected to by the Examiner.								
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).								
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d). 11) The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.								
•—	der 35 U.S.C. § 119	•						
12) Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f). a) All b) Some * c) None of: 1. Certified copies of the priority documents have been received. 2. Certified copies of the priority documents have been received in Application No 3. Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)). * See the attached detailed Office action for a list of the certified copies not received.								
2) Notice 3) Informa	of References Cited (PTO-892) of Draftsperson's Patent Drawing Review (ation Disclosure Statement(s) (PTO-1449 o		4) Interview Summa Paper No(s)/Mail 5) Notice of Informa 6) Other:					

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DETAILED ACTION

The applicant's response filed 9/5/2004 to the Office action has been entered. Claims 1-68 are pending. Claims 51-67 are withdrawn from consideration.

- 1. The rejection of claim 49 under 35 U.S.C. 112, second paragraph is withdrawn.
- 2. Claims 1-25 and 68 remain rejected under 35 U.S.C. 103(a) as being unpatentable over Pritham et al. (J of Clinical Ligand Assay, 1998, Vol. (4), pg. 404-412) in view of Johnston-Dow et al. (6,103,465).

Pritham et al. disclose a rapid PCR method to monitor the amplification by detecting the fluorescent signal (See pg. 404, the abstract) involving using fluorescence probe (See pg. 405 column 2, second paragraph and pg 406 column 2 to pg. 409, column 1). The teachings of Pritham et al. are recited through out the limitations of claims 1-9, and 19-24, except that Pritham et al do not disclose the sequencing method used to detect a specific target nucleic acid as recited in the limitations of claim 1.

Pritham et al. also do not indicate the source of the DNA sample used as listed in claims 10, and 25 in the method.

Johnston-Dow et al. disclose a method for typing HLA class I gene and the method involving DNA sequencing techniques (See the Abstract and column 9, lines 9-22). The method is to provide for the specific DNA sequencing of HLA-A, HLA-B and HLA-C (See column 3, lines 19-22). Johnston-Dow et al. also disclose that any source of human nucleic acid can be used, for example, blood and lymphoblostoid cell lines (See column 6, lines 9-14) as recited in the limitations of claims 10, and 25. Johnston-Dow et al. further indicate that HLA typing is performed routinely in connection with many medical indications, the study of auto-immune disease and the determination of susceptibility to infectious disease (See column 1, lines 57-62). This teaching suggests the limitations of claims 11-18 in that the pathogen will be from a virus,

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prokaryote and eukaryote, the presence of the given target polynucleotide indicates the presence of the genetic disease or a specific allele which can indicate serotype.

It would have been prima facie obvious to an ordinary skill in the art at the time of the instant invention to combine the teachings of Pritham et al. and Johnston-Dow et al. to carry out the method as claimed with a reasonable expectation of success. The motivation is that the teachings of Pritham et al. indicate that fluorescent monitoring of PCR provides qualitative and quantitative information in that the qualitative information includes purity and identity (See pg. 404, column 1, last paragraph) and rapid cycle PCR is an ideal technique for fluorescence monitoring because temperature gradients within samples are minimized (See pg. 404, column 2, second paragraph) and the method of Johnson-Dow et al. is applied to the locus-specific nucleic acid amplification followed by sequence-specific detection of the amplified product for the DNA typing of HLA class I gene via DNA sequencing in that by sequencing the exons in both directions, the effect of sequencing errors on the assignment of HLA type is minimized and the method greatly reduces the number of reagents and the complexity of the sequencing protocols required (See column 9, lines 29-37).

The response argues that the instant invention has amplified products that is directly sequenced is not subject to "meticulous purification" prior to the sequencing and thus the applied art would not provide a reasonable expectation of success. However, the reference of Johnston-Dow indicates that sequencing was performed with no purification of the PCR product using the TaqCS polymerase enzyme (See column 13, lines 39-40 and column 18, lines 36-58). Thus, the rejection is maintained.

3. Claims 26-50 remain rejected under 35 U.S.C. 103(a) as being unpatentable over Pritham et al. (J of Clinical Ligand Assay, 1998, Vol. (4), pg. 404-412) in view of Johnston-Dow et al.

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(6,103,465) as applied to claims 1-25 and 68 above, and further in view of Wittwer et al. (6,174,670).

The teachings of Pritham et al. and Johnston-Dow et al. are set forth in section 2 above. The teachings of Pritham et al. and Johnston-Dow et al. do not indicate that there are two reaction compositions involved in the methods.

Wittwer et al. disclose methods of monitoring hybridization during polymerase chain reaction using two pairs of oligonucleotides and a nucleic acid binding fluorescent dye to monitor amplification of a selected template (See column 13, lines 62 to column 14, lines 29).

Thus, it would have been <u>prima facie</u> obvious to an ordinary skill in the art at the time of the instant invention to combine the teachings of Pritham et al., Johnston-Dow et al. and Wittwer et al. to carry out the method as claimed with a reasonable expectation of success. The motivation of combining the teachings of Pritham et al. and Johnston-Dow et al. are discussed in section 5 above and the motivation of applying the teachings of Wittwer et al. is that the method of Wittwer et al. improves the sensitivity of PCR quantification and reduces the time of fluorescence monitoring for PCR.

The response argues that the claim language is incorrectly interpreted. However, the language "combining nucleic acid from the sample with at least one set of reaction composition comprising a first reaction composition and second reaction composition, ..." can be interpreted as that the nucleic acid is combined with two compositions in one reaction or the nucleic acid is combined with first composition in one reaction and the nucleic acid is combined with second composition in another reaction.

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The response further argues that the claim language clearly present there are two reactions because "... the first reaction composition comprising amplification primers specific to the at least one target polynucleotide, and the second reaction composition comprises a fluorescent indicator and amplification primer specific to the at least one target polynucleotide". Nevertheless, "the at least one target" can be interpreted as the same target polynucleotide in each of two reactions or the different polynucleotide targets in one reaction. Thus, the language can be interpreted that the reaction is either one reaction or two reactions.

Moreover, the response cites the disclosure from the specification (See pg. 14, paragraph 42). However, the limitations are read in light of the specification, but the limitations have to be in the claims.

Regardless how the claim language is interpreted, the teachings of Wittwer et al. still read on the limitations of the claims (See column 13, lines 63-76 and column 14, lines 1-43).

Therefore, the rejection is maintained.

Summary

- 4. No claims are allowable.
- 5. THIS ACTION IS MADE FINAL. Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period

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will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event however, will the statutory period for reply expire later than SIX MONTHS from the mailing date of this final action.

6. Any inquiries concerning this communication or earlier communications from the examiner should be directed to Joyce Tung whose telephone number is (703) 305-7112. The examiner can normally be reached on Monday-Friday from 8:00 AM-4:30 PM.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Gary Benzion can be reached at (703) 308-1119 on Monday-Friday from 10:00 AM-6:00 PM.

Any inquiries of a general nature or relating to the status of this application should be directed to the Chemical/Matrix receptionist whose telephone number is (703) 308-0196.

7. Papers related to this application may be submitted to Group 1600 by facsimile transmission. Papers should be faxed to Art Unit 1637 via the PTO Fax Center located in Crystal Mall 1 using (703) 305-3014 or 308-4242. The faxing of such papers must conform with the notice published in the Official Gazette, 1096 OG 30 (November 15, 1989).

Joyce Tung TT November 19, 2004

SUPERVISORY PATENT EXAMINER

TECHNOLOGY CENTER 1600